

US EPA ARCHIVE DOCUMENT



# PROPOSAL

## Interim Guidance for the Efficacy Evaluation of Products with Biofilm Claims

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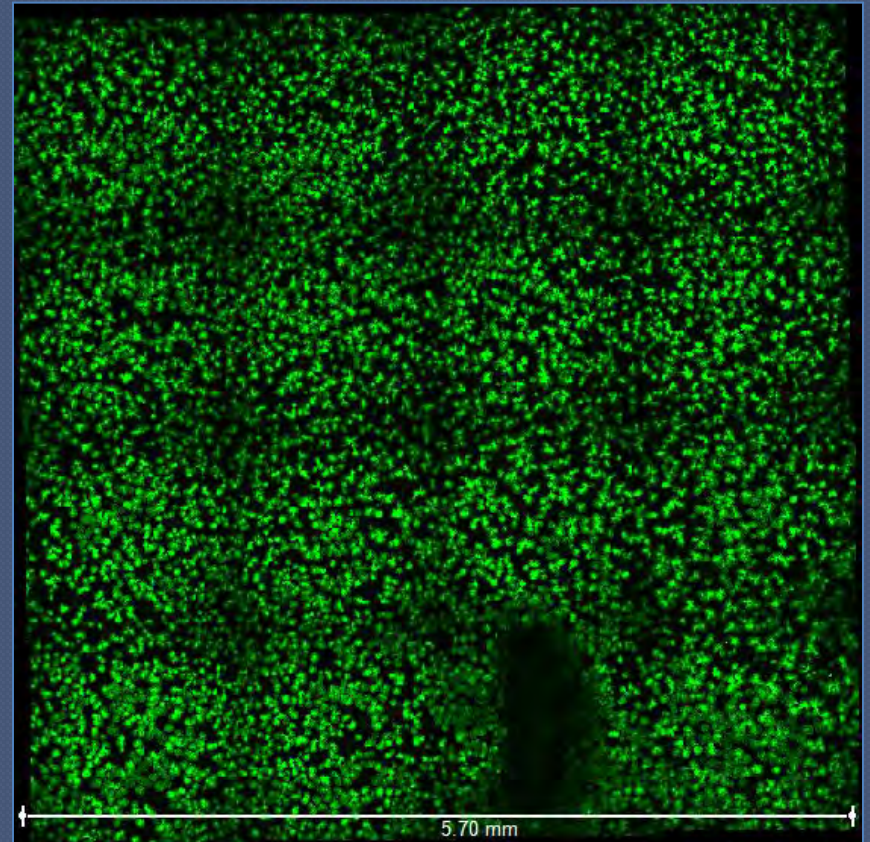
# Disclaimer

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# Background

- Biofilm is considered to be a “pest” by the EPA.
- Therefore, any label claim to prevent, destroy, repel or mitigate biofilm on an inanimate environmental surface is a pesticidal claim which requires registration under FIFRA – including product efficacy data.
- Biofilm express unique characteristics, and therefore require unique and relevant test methods for measuring product efficacy.
- The choice of method will dictate the type of label claim.



# Scope

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- Agency's proposed interim guidance for the efficacy evaluation of antimicrobial pesticides sold as water soluble powders or liquids that are labeled for treating hard non-porous surfaces contaminated with bacterial biofilm.
- Registrants and applicants may propose and submit alternative practices to the Agency for assessment and the Agency will evaluate them for appropriateness on a case-by-case basis.
- This guidance may be updated in the future.



# Product Efficacy

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- ◉ The following quantitative test method may be used to generate efficacy data:
  - ASTM E2871-12: Standard Test Method for Evaluating Disinfectant Efficacy against *Pseudomonas aeruginosa* Biofilm Grown in CDC Biofilm Reactor using Single Tube Method.
- ◉ A claim for controlling biofilm may be added to a registered antimicrobial product with an existing claim.

# Example 1

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- ◉ For a limited-spectrum disinfectant claim, the product must pass all of the following efficacy tests:
  - AOAC Use-Dilution Method for *Staphylococcus aureus* (AOAC 955.15) or *Salmonella enterica* (AOAC 955.14) and the
  - ASTM Single Tube Method (ASTM E2871-12) for *P. aeruginosa*

## Example 2

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- For a broad-spectrum disinfectant claim, the product must pass all of the following efficacy tests:
  - AOAC Use-Dilution Method for *S. aureus* (AOAC 955.15)
  - AOAC Use-Dilution Method for *S. enterica* (AOAC 955.14) and the
  - ASTM Single Tube Method (ASTM E2871-12) for *P. aeruginosa*



## Example 3

- For a hospital or healthcare disinfectant claim, the product must pass all of the following efficacy tests:
  - AOAC Use-Dilution Method for *P. aeruginosa* (AOAC 964.02)
  - AOAC Use-Dilution Method for *S. aureus* (AOAC 955.15) and the
  - ASTM Single Tube Method (ASTM E2871-12) for *P. aeruginosa*
  - ASTM Single Tube Method (ASTM E2871-12) modified for *S. aureus*

# Options

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- Applicants should consult with the Agency for other formulations prior to developing registration data. The Agency may require submissions of protocols for review.
- Modifications to the test method will be necessary to accommodate other biofilm forming microbes.

# Test Strains/Test Cultures

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## ◉ *Pseudomonas aeruginosa*

- Produce stock cultures and a mature biofilm of *P. aeruginosa* according to the EPA's Microbiology Laboratory Branch's (MLB) standard operating procedure (SOP) MB-19-02.
- Note: *P. aeruginosa* (ATCC 15442) may display three colony types.

# Test Strains/Test Cultures

## ◉ *Staphylococcus aureus*

- ASTM standards do not currently address the culture of *S. aureus*.
- Produce a mature *S. aureus* biofilm per EPA MLB SOP MB-19-02 with the following exceptions:
  - *S. aureus* ATCC 6538 is utilized.
  - Biofilm cultures are initiated using a frozen stock culture of *S. aureus* (refer to Attachment 2 of EPA MLB SOP MB-05-11 for preparation of frozen stock cultures)
  - Use tryptic soy broth (30 g/L).
  - Incubate batch phase at  $36 \pm 1^\circ\text{C}$  with a baffle speed of 60 rpm.
  - Use tryptic soy broth (300 mg/L) as the growth medium for continuous flow operation (CSTR) mode.

# Test Parameters

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## ● Product Performance

- A minimum 5 log reduction in viable bacteria in biofilm is required.
- The mean log density for carriers inoculated with *P. aeruginosa* or *S. aureus* must be at least 8.0 (corresponding to a geometric mean density of  $1.0 \times 10^8$ ).

## ● Contact Time

- The contact time for testing should not exceed 10 minutes.



# Other Specifics

- Carrier Material

- Borosilicate glass coupons

- Number of Batches

- Test three batches at the lower certified limit(s) (LCL)

- Test Carriers per Batch

- Evaluate a minimum of five carriers against the disinfectant and 3 carriers as controls.

- Neutralizer Confirmation

- Conduct neutralization testing in advance of efficacy testing to determine the appropriate neutralizer for the product.
  - Use MLB SOP MB-20-01 Attachment 1.
  - Submit neutralizer confirmation data along with product efficacy data.

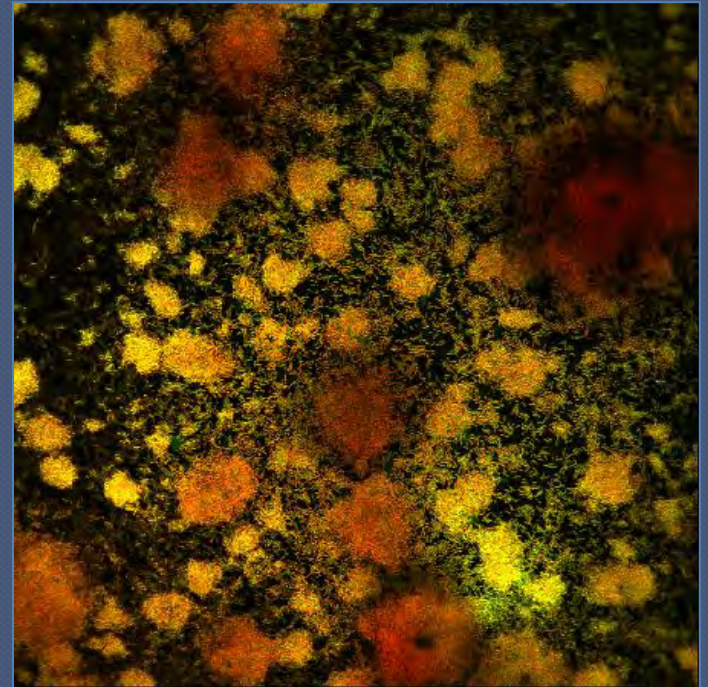
# Label Claims

## Examples of Claims:

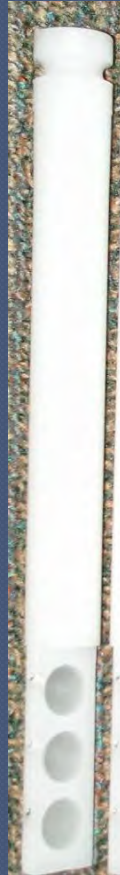
- Kills 99.999% of bacteria in biofilm on a hard, non-porous surface
- Kills a minimum of 99.999% of bacteria in biofilm
- Reduces at least 99.999% of bacteria growing in biofilm

## Other related claims:

- Kills biofilm bacteria
- Controls slime-forming bacteria
- Specifically designed and formulated to destroy bacteria in biofilm
- Penetrates biofilm, killing the bacteria living there

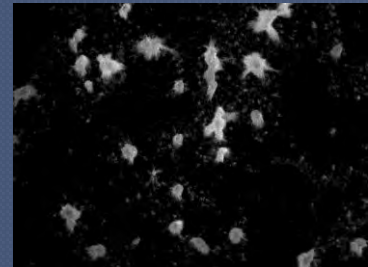


# CDC Biofilm Reactor

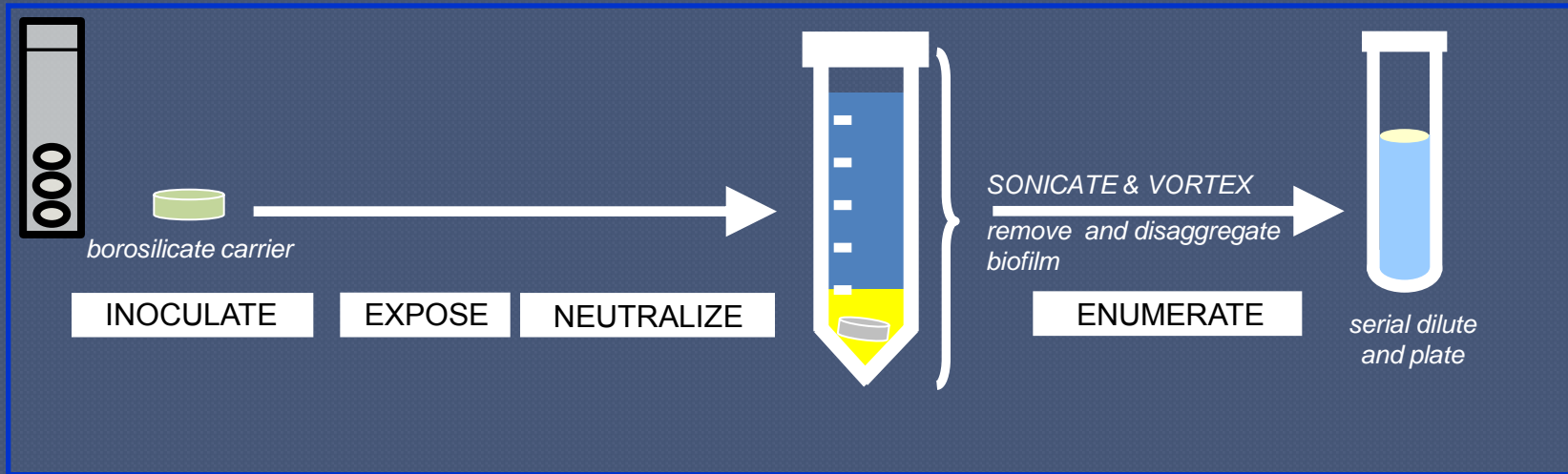


Eight Rods per reactor

Three coupons per rod



# Single Tube Method



## Key Aspects/Steps:

- 4 mL disinfectant
- 36 mL neutralizer
- Variable contact times & concentrations
- $20 \pm 1^\circ\text{C}$  treatment temperature

- Vortex 30 s
- Sonicate 30 s
- Repeat twice
- Vortex 30 s

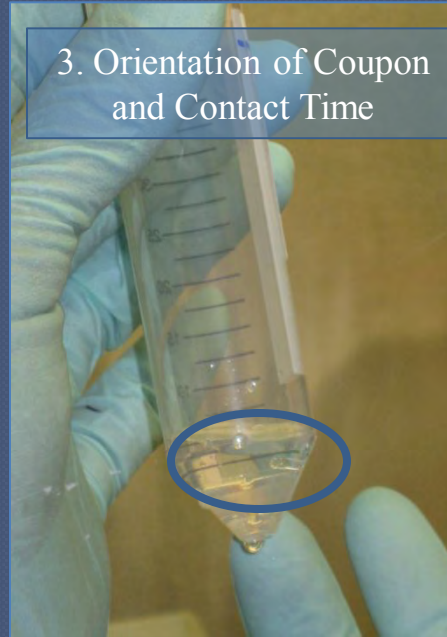


# Single Tube Method

1. Extraction of Coupons



3. Orientation of Coupon and Contact Time



4. Addition of Neutralizer to Stop Activity



2. Addition of Disinfectant



5. Sonication and Vortex



6. Dilute and Plate





## Next Steps

- ◉ Propose revisions to the ASTM methods; especially for testing of *S. aureus* biofilm or make revisions to MLB's SOPs
- ◉ To increase accuracy of the assay:
  - Filter the entire contents of reaction tube containing the carrier
  - Data generated from filtration can be used to assess LRs as high as the control counts in the event of complete kill.
- ◉ Pre-screening test chemicals may be necessary to determine the appropriate dilutions required to achieve countable plates
- ◉ Work with MSU on method performance criteria
- ◉ Finalize guidance and post by September 2014

# Acknowledgements

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